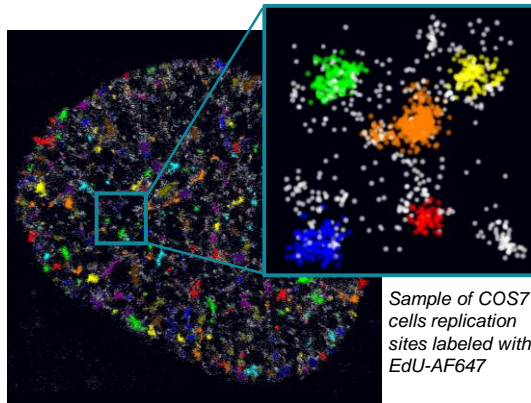


How to analyze clusters in SMLM data

Single-molecule localization microscopy is particularly adapted for clustering analysis, considering that the final dataset is not simply an image, but the coordinates of individual molecules. Many applications require clustering analysis: clathrins, nuclear pores, bacteria, DNA replication sites... biological entities tend to accumulate and form 3D clusters whose size, density and distribution are worth quantifying. These factors can then describe qualitatively a biological structure, and they can be compared quantitatively between conditions, for example to measure the effect of a given drug on the size of clusters, etc. Here, we describe the main algorithms developed to analyze clusters of single molecules.



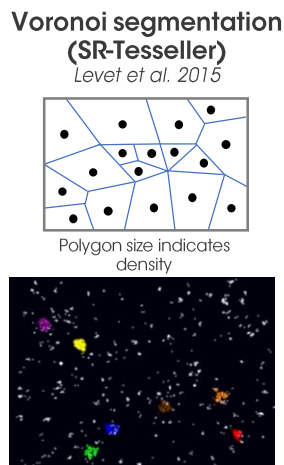
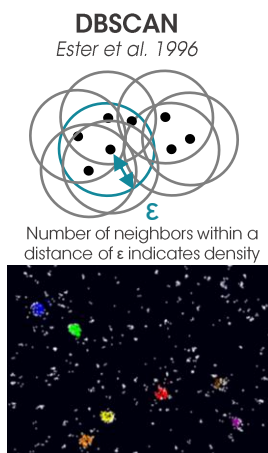
Sample of COS7 cells replication sites labeled with EdU-AF647

How can I isolate individual clusters ?

Several algorithms enable the identification of clusters in a dataset, including:

DBSCAN (Density Based Spatial Clustering of Applications with Noise) requires two input parameters: a distance ϵ and a minimum number of neighbors MinPts. For each localization in the dataset, the algorithm searches whether it has enough neighbors MinPts within the distance ϵ . If yes, it considers the localization as part of a cluster, etc.

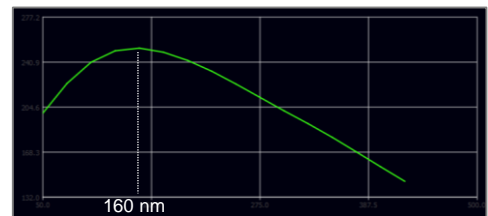
Voronoi partitions the image into polygons, where each polygon contains one, and only one, localization. The area/volume of the polygon is indicative of the density of localizations: a dense region will have small polygons while a low-density region will have big polygons. The user can choose a density threshold, above which localizations are considered as part of a cluster.



In practice

How do I know if there are clusters in my dataset?

The K Ripley function evaluates whether a population of localizations is aggregated or randomly distributed based on a neighborhood analysis.



The bell-shaped curve indicates the presence of aggregated datapoints and provides an estimate of size of these aggregates.

Abbelight SFAe modules and NEO software offer features covering:

- Descriptive spatial statistics
- Optimized clustering algorithms using smart and fast spatial partitioning of data:
 - K Ripley function
 - DBSCAN algorithm
 - Voronoi segmentation
- Quantitative cluster info: number of clusters, cluster surface area/volume, density number of localizations, centroids, radius of gyration
- 3D real-time interactive visualization
- Export of quantitative results

Quantification

Clusters	1	2	3
Centroid			
X	669	654	678
Y	526	514	525
Z	0.313	0.302	0.308
(μm)			
Number of localizations	277	249	256
Volume (μm^3)	0.015	0.019	0.017

